## **Promega**

## ReliaPrep™ miRNA Cell and Tissue Miniprep System

Instructions for Use of Products Z6210, Z6211 and Z6212.

Quick Protocol

## **RNA Isolation and Purification Procedure from Tissue Samples**

Use the following protocol to lyse tissue samples. Use 0.25-20mg of tissue per purification.

1. Place fresh or flash-frozen tissue samples in a sterile centrifuge tube containing 200µl LBA + TG. Use a mechanical homogenizer or mini pestle to disrupt the tissue until homogeneous.

**Note:** Following lysis, pipet 7–10 times to shear the DNA using a P200 or P1000 pipettor.

- 2. Add 130 $\mu$ l of RDB to each homogenate and vortex for 10 seconds. Centrifuge for 2 minutes at 12,000  $\times$  g. Carefully transfer the cleared homogenate to a clean 1.5ml tube.
- 3. Add 400µl of 100% isopropanol to each cleared homogenate. Mix by vortexing.
- 4. Transfer the homogenate to a ReliaPrep<sup>TM</sup> Minicolumn. Centrifuge at  $12,000 \times g$  for 30 seconds.
- 5. Remove the column and discard the liquid. Place the column back into the Collection Tube.
- 6. Transfer the remaining homogenate liquid onto the same column used in Step 5. Centrifuge at  $12,000 \times g$  for 30 seconds.
- 7. Remove the column and discard the liquid. Place the column back into the Collection Tube.
- 8. Add 500µl of RWA to each column. Centrifuge at 12,000  $\times$  g for 30 seconds.
- 9. Remove the column and discard the liquid. Place the column back into the Collection Tube.
- 10. Add 500 $\mu$ l of RWA to each column. Centrifuge at 12,000  $\times$  g for 2 minutes. Carefully transfer the column to a 1.5ml Elution Tube.
- 11. Add 40µl of Nuclease-Free Water to each column. Centrifuge at 12,000  $\times$  g for 1 minute.
- 12. Transfer 5µl of DNase 10X Buffer and 5µl of DNase I to eluate.
- 13. Incubate for 5 minutes at room temperature (20–25°C).
- 14. Add 150µl of LBA + TG Buffer to the DNase treatment tube.
- 15. Add 300 $\mu$ l of 95% ethanol to the mixture and vortex for 10 seconds. Transfer 500 $\mu$ l of this mixture to a new column. Centrifuge at 12,000  $\times$  g for 30 seconds.
- 16. Remove the column and discard the liquid. Place the column back into the Collection Tube and repeat Steps 8–10.
- 17. Add 15 $\mu$ l of Nuclease-Free Water to each column (see Table 1). Centrifuge at 12,000  $\times$  g for 1 minute.

Table 1. Recommended RNA Elution Volumes per Milligram of Tissue.

Tissue Input Range	Nuclease-Free Water
0.25-5mg	15µl
>5-10mg	30µІ
>10mg	50µІ

**Note:** RNA concentration may increase with lower elution volumes; however, the total yield of RNA may decrease when elution volumes are between  $10-15\mu$ l. If maximum recovery of RNA is essential, we recommend a second elution into a second sterile tube with an additional  $15\mu$ l of Nuclease-Free Water followed by centrifugation at  $12,000 \times g$  for 1 minute.



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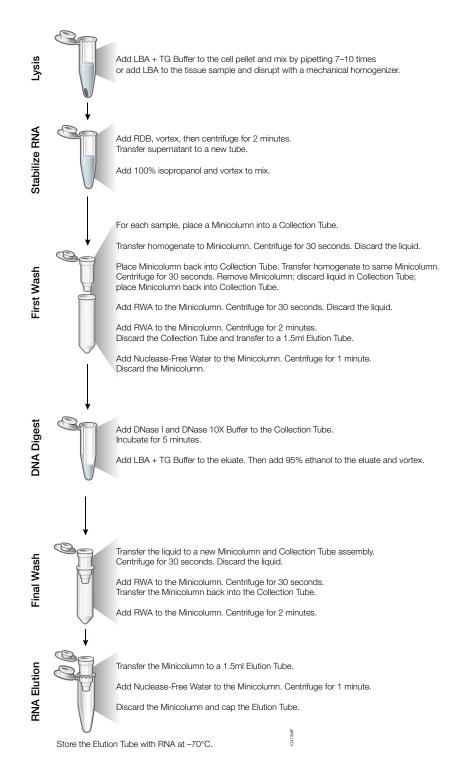


Figure 1. Schematic diagram of the ReliaPrep™ miRNA Cell and Tissue Miniprep System.

Additional protocol information is in Technical Manual #TM469, available online at: www.promega.com

